

Introduction

Lentiviral Packaging Kit is designed to optimize the packaging of third generation lentivectors into virus particles which can express your lentiviral construct in a multitude of mammalian cells. The 3rd generation packaging system offers maximal biosafety as the lentiviral Rev gene is supplied as an independent vector from other structure genes, further eliminating the possibility of reverse recombination of vectors into replication competent viral particles.

Package Information

| Components | M0040 |
|--------------------------------|--------------------|
| Package Plasmid Mix | 50 µl (0.5 µg/µl) |
| Control Plasmid | 50 µl (0.25 µg/µl) |
| LiFect2K™ Transfection Reagent | 250 µl |

Storage

Package plasmid mix and control plasmid at -20°C; LiFect2K™ reagent at 4°C

Additional Materials Required

- HEK293T cell line (ATCC Cat #: CRL-11268)
- H1299 cell line (ATCC Cat #: CRL-5803)
- DMEM media (Mediatech Cat #: 10-013-CV)
- Fetal bovine serum (Biosera Cat #: FB-1001/500)
- Opti-MEM I Reduced-Serum Medium (Thermo Fisher Cat #: 31985062)
- Polybrene (Cat #: M0091), Lentivirus Transduction Enhancer I (Cat #: M0089) or Lentivirus Transduction Enhancer II (Cat #: M0090)
- Penicillin-Streptomycin (Thermo Fisher Cat #: 15070063)
- BD Falcon® 5ml or 14 ml tubes (BD Falcon® Cat #: 352053 & 352059)

Lentivirus Production Protocol

The following protocol provides optimized steps for lentivirus production in 293T packaging cells. The yield of recombinant lentiviral particles typically produced under these optimized conditions is 10 ml of 1-10 x10⁶ infection units (ifu) per ml of un-concentrated supernatant from one 10-cm culture dish for eGFP positive controls. This amount of pseudoviral particles is generally sufficient to infect 1-10 x10⁷ target cells at a MOI (multiplicity of infection) equal to 1. The titers of lentivirus decrease as the size of insert increases. Actual lentivirus titers for your gene

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Cat. #: M0040 Size: 10 rxns

of interest will vary accordingly.

Caution: Following this protocol results in the production of pseudoviral particles capable of infecting mammalian cells. The recommended guidelines for working with BSL-2 safety class must be adhered to.

1. Plate packaging cells

Two days before transfection, plate 1.3-1.5 x10⁶ of HEK293T cells in a 10cm dish in 10 ml of DMEM supplemented with 10% heat-inactivated fetal bovine serum so that the cells are 70-80% confluent at the moment of transfection. Incubate the cells at 37°C with 5% CO₂.

Note: Plating the HEK293T cells 2 days prior to transfection significantly increases the titer of lentivirus. Use heat-inactivated fetal bovine serum for lentivirus production.

2. Prepare DNA/transfection reagent complex

In a sterile polypropylene tube, dilute 2.5 µg of lentiviral ORF/shRNA/miRNA expression plasmid or control plasmid, and 5.0 µl (0.5 µg/µl) of Package Plasmid Mix into 200 µl of Opti-MEM® I Medium. In a separate tube, dilute 15 µl of LiFect2K™ Transfection Reagent into 200 µl of Opti-MEM® I Medium. Add diluted LiFect2K™ Transfection reagent dropwise to the DNA solution while gently vortexing the DNA-containing tube. Incubate the mixture for 20 minutes at room temperature.

Note: The DNA-LiFect2K™ complex must be formed in the absence of proteins even though the complex is able to transfect cells in the presence of proteins such as 10% serum. Opti-MEM® I Medium is recommended for diluting both DNA and LiFect2K™ Transfection reagent. Serum-free DMEM can be used in place of Opti-MEM® I Medium but the transfection efficiency will be compromised.

3. Transfect packaging cells

Add the DNA-LiFect2K™ complex directly to each dish and gently swirl the dish to distribute the complex. Incubate the cells in a CO₂ incubator at 37°C overnight (8-14 hours). Replace the overnight culture medium with fresh DMEM medium supplemented with 2-5% heat-inactivated fetal bovine serum and penicillin-streptomycin, and continue incubation in the CO₂ incubator at 37°C.

4. Harvest lentivirus

After 48 hours post transfection, collect the pseudovirus-containing culture medium in sterile capped tubes and centrifuge the tubes at 500× g for 10 minutes to get rid of cell debris. Following centrifugation, filter the supernatant through 0.45 µm polyethersulfone (PES) low protein-binding filters.

Note: If you want to collect more lentivirus, replace lentiviral containing supernatants with fresh DMEM medium supplemented with 2-5% heat-inactivated fetal bovine serum,



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penicillin-streptomycin. Lentivirus containing medium can be collected again at 72 hours post-transfection. Do not use nitrocellulose filters as nitrocellulose is known to bind lentivirus and reduce titers.

The supernatant containing lentiviral particles can be used directly to determine the titer and to transduce target cells in vitro as long as the target cells can survive in conditioned medium. Lentiviral stocks should be aliquoted and stored at -80°C . Expect significant loss of viral titer with each freeze/thaw cycle.

Lentivirus Titration

The titer of harvested lentiviral stocks can be determined by commercial qPCR Lentiviral Titration Kits.

Transduction of Target Cells with Lentiviruses

The transduction efficiency of mammalian cells varies significantly under different experimental conditions. This includes virus concentration, exposure time to the virus and growth area of the well or plate used for the infection.

To determine the viral concentration required to provide the desired multiplicity of infection (MOI) for your target cells, it is advisable to perform several test transductions with reporter viral particles at a range of different volumes such as 1 μl , 5 μl , 10 μl , and 100 μl . Results from these preliminary tests can be used to determine an optimal concentration that will yield the highest percentage of successfully infected cells.

The following protocol has been provided as a general guideline only, to be used as a starting point for determining optimal conditions for target cell transduction:

1. Plate the target cells in a 24-well plate, 24 hours prior to viral infection at a density of 5×10^4 cells per well. Add 0.5 ml of complete optimal medium (with serum and antibiotics if required) and incubate the cells at 37°C with 5% CO_2 overnight.
2. For each well, prepare 0.5 ml of virus suspension diluted in complete medium with polybrene at a final concentration of 8 $\mu\text{g}/\text{ml}$.

Note: Use several dilutions of pseudoviral stock (0.1 μl to 100 μl). We recommend gradient dilution of 0.1 μl , 0.3 μl , 3 μl , 10 μl , 30 μl for standard particles, and 0.1 μl , 0.3 μl , 0.5 μl , 0.7 μl , 0.9 μl for purified particles. Mix the virus with the medium gently by inverting the tubes several times. Do not vortex.

3. Infect the target cells by removing the old culture medium and replacing it with 0.5 ml of diluted viral supernatant. Place the plates in a 37°C incubator with 5% CO_2 and incubate cells overnight. If the transduction efficiency of the target cells is low, try Lentivirus Transduction Enhancer I (Cat #: M0089), or Lentivirus Transduction Enhancer II (Cat #: M0090).

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4. Replace the old medium with 0.5 ml of fresh complete medium (without Polybrene).
5. The infected target cells can be analyzed for transient expression of transgenes using an appropriate biological assay. If you have used an internal eGFP control, determine the percentage of infected cells with a fluorescent microscope.
6. Once an effective MOI has been determined for the target cells through preliminary test infections, use the appropriate volume of virus to infect your cells.